

rpc00086

Nicotiana tabacum* NaCl-r cell suspension culture*Components**

- Domestic delivery: A 50-mL plastic conical centrifuge tube, containing cell suspension
- Overseas delivery: A 250-mL plastic Erlenmeyer flask, containing cells placed on semi-solid medium

Notice

- Subculture the cells to fresh medium immediately after arrival [[Notes I](#), [II](#)].
- Do not store the cell culture in a refrigerator and a freezer.
- Maintain aseptic conditions of the cell culture, and work in a laminar flow cabinet.

Method

- Culture medium: mLS medium, 10 μ M NAA, 1 μ M kinetin, 0.2 M NaCl, pH 5.7 (medium no. 56) [[Materials III](#)]
- Culture conditions: 24°C, continuous light, 90 rpm [[Methods II](#)]
- Subculture: 21-day intervals [[Methods I](#)]

Citation of cell line

When results obtained by using this cell line are published in a scientific journal, it should be cited in the following manner: “*Nicotiana tabacum* NaCl-r cell line (rpc00086) was provided by the RIKEN BRC through the National BioResource Project of the MEXT, Japan.”

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Introduction

Tobacco NaCl-r cell line is a NaCl-adapted NI cell line, which can grow photoautotrophically in a NaCl-containing medium (Murota *et al.* 1994). The parent photoautotrophic NI cell line (rpc00084) was established from a pith of *Nicotiana tabacum* L. cultivar Samsun NN (Yamada and Sato 1978). The NaCl-r cells are grown in a modified Linsmaier and Skoog (mLS) medium supplemented with 10 μ M 1-naphthaleneacetic acid (NAA), 1 μ M kinetin and 0.2 M NaCl, pH 5.7. Our NaCl-r cell culture has been maintained under the continuous light at 24°C with rotary shaking at 90 rpm and subcultured at 21-day intervals.

Materials

Chemicals and stock solutions

(All stock solutions are stored at 4°C)

A) MS salt mix

Murashige and Skoog Plant Salt Mixture, FUJIFILM Wako Pure Chemical Corporation (#392-00591)

B) Sucrose

C) LS_VT

Thiamine·HCl	0.16 mg/mL
<i>myo</i> -Inositol	40 mg/mL

D) NAA (1 mM)

NAA·K	0.224 mg/mL
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Potassium 1-naphthylacetate, FUJIFILM Wako Pure Chemical Corporation (#161-04021)

E) Kinetin (1 mM)

Kinetin	0.215 mg/mL
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Dissolve kinetin in small volume of KOH (1 N), and fill up with distilled water

F) NaCl

G) KOH (1 N)

Glassware

A) Erlenmeyer flask (100 mL), capped with two layers of aluminum foil

B) Pipette (10 mL; large tip opening) and a bulb, sterilized by autoclaving at 121°C for 20 min

Preparation of mLS medium (medium no. 56)

1. Dissolve the following chemicals in approximately 800 mL of distilled water.

MS salt mix	1 bag (1 L)
Sucrose	30 g
NaCl	11.69 g

2. Add following stock solutions, and fill up to approximately 950 mL with distilled water.

LS_VT	5 mL
NAA (1 mM)	10 mL
Kinetin (1 mM)	1 mL

3. Adjust the pH of the solution to 5.7 with KOH (1 N), and fill up to 1 L with distilled water.
4. Pour 25 mL of the medium into a 100-mL flask.
5. Autoclave the flask at 121°C for 20 min.

Methods

1. Agitate a 21-day-old culture well and transfer 5–10 mL of cell suspension to 25 mL of fresh mLS medium with a pipette.
2. Incubate cell cultures on a rotary shaker at 90 rpm under the continuous light condition (photosynthetic photon flux density 50–60 $\mu\text{mol m}^{-2} \text{s}^{-1}$) at 24°C.

Notes

- For domestic customers: We send NaCl-r cell suspension in a 50-mL disposable conical centrifuge tube. The cells should be transferred to fresh mLS medium immediately after arrival. Transfer settled cells to Erlenmeyer flasks containing fresh liquid medium with a pipette.
- For overseas customers: We send NaCl-r cells placed on semi-solid mLS medium in a 250-mL disposable Erlenmeyer flask. The cells should be transferred to fresh mLS medium immediately after arrival. Collect the cells from the semi-solid medium with a spatula and transfer them to Erlenmeyer flasks containing fresh liquid medium.
- In order to maintain NaCl-r cell suspension cultures stably, it is essential to transfer an adequate amount of cells to fresh mLS medium in every subculture. The amount of cells may vary from one lab to another, because proliferation of NaCl-r cells is

affected by culture conditions, such as a room temperature, rotation speed of a rotary shaker, and aeration condition of the culture.

- In order to obtain good aeration of a suspension culture, a silicone sponge plug may be used instead of the aluminum foil cap (*e.g.*, cap-type Silicosen; Shin-Etsu Polymer, Tokyo, Japan; <https://www.shinpoly.co.jp/en/product/product/medical/plugs.html>).

References

- Murota K, Ohshita Y, Watanabe A, Aso S, Sato F, Yamada Y (1994) Changes related to salt tolerance in thylakoid membranes of photoautotrophically cultured green tobacco cells. *Plant & Cell Physiology* 35: 107–113. DOI: [10.1093/oxfordjournals.pcp.a078560](https://doi.org/10.1093/oxfordjournals.pcp.a078560)
- Yamada Y, Sato F (1978) The photoautotrophic culture of chlorophyllous cells. *Plant & Cell Physiology* 19: 691–699. DOI: [10.1093/oxfordjournals.pcp.a075640](https://doi.org/10.1093/oxfordjournals.pcp.a075640)

Appendix A: Formulation of culture medium

Table A1. modified Linsmaier and Skoog medium
(medium no. 56)

Chemical	Concentration (mg/L)
KNO ₃	1900
NH ₄ NO ₃	1650
CaCl ₂ ·2H ₂ O	440
MgSO ₄ ·7H ₂ O	370
KH ₂ PO ₄	170
H ₃ BO ₃	6.2
MnSO ₄ ·4H ₂ O	22.3
ZnSO ₄ ·7H ₂ O	8.6
KI	0.83
Na ₂ MoO ₄ ·2H ₂ O	0.25
CuSO ₄ ·5H ₂ O	0.025
CoCl ₂ ·6H ₂ O	0.025
FeSO ₄ ·7H ₂ O	27.8
Na ₂ -EDTA	37.3
Thiamine·HCl	0.8
<i>myo</i> -Inositol	200
Sucrose	30000
NAA·K	2.24
Kinetin	0.215
NaCl	11.69