

rpc00040

Nicotiana tabacum* GF11 transgenic cell suspension culture*Components**

- Domestic delivery: A 50-mL plastic conical centrifuge tube, containing cell suspension
- Overseas delivery: A 250-mL plastic Erlenmeyer flask, containing cells placed on semi-solid medium

Notice

- Subculture the cells to fresh medium immediately after arrival [[Notes I](#), [II](#)].
- Do not store the cell culture in a refrigerator and a freezer.
- Maintain aseptic conditions of the cell culture, and work in a laminar flow cabinet.

Method

- Culture medium: mLS medium, 0.2 mg/L 2,4-D, pH 5.8 (medium no. 1) [[Materials III](#)]
- Culture conditions: 27°C, dark, 130 rpm [[Methods II](#)]
- Subculture: 7-day intervals [[Methods I](#)]

Citation of cell line

When results obtained by using this cell line are published in a scientific journal, it should be cited in the following manner: “*Nicotiana tabacum* GF11 cell line (rpc00040) was provided by the RIKEN BRC through the National BioResource Project of the MEXT, Japan.”

Experimental Plant Division
RIKEN BioResource Research Center (BRC)
Koyadai 3-1-1, Tsukuba, Ibaraki 305-0074
Japan
FAX: +81 29 836 9053
E-mail: plant.brc@riken.jp
<http://epd.brc.riken.jp/en/>

Introduction

Tobacco GF11 cell line is a transgenic BY-2 cell line expressing Green Fluorescent Protein (GFP) fused with Arabidopsis fimbrin second actin-binding domain (ABD2) (Sano *et al.* 2005). GFP fluorescence is observed in actin microfilaments by using a fluorescence microscope. The parent cell line BY-2 (rpc00001) was established from a callus induced from a seedling of *Nicotiana tabacum* L. cultivar Bright Yellow 2 (Nagata *et al.* 1992). The GF11 cells are grown in a modified Linsmaier and Skoog (mLS) medium supplemented with 0.2 mg/L 2,4-dichlorophenoxyacetic acid (2,4-D), pH 5.8. Our GF11 cell culture has been maintained in the dark at 27°C with rotary shaking at 130 rpm and subcultured at 7-day intervals.

Materials

Chemicals and stock solutions

(All stock solutions are stored at 4°C)

A) MS salt mix

Murashige and Skoog Plant Salt Mixture, FUJIFILM Wako Pure Chemical Corporation (#392-00591)

B) Sucrose

C) BY2_P

KH_2PO_4 80 mg/mL

D) LS_VT_modified

Thiamine·HCl 0.4 mg/mL
myo-Inositol 40 mg/mL

E) 2,4-D (0.2 mg/mL)

2,4-D sodium monohydrate 0.236 mg/mL
(2,4-Dichlorophenoxy)acetic acid sodium salt monohydrate, Sigma-Aldrich (D6679)

F) KOH (1 N)

Glassware

A) Erlenmeyer flask (300 mL), capped with two layers of aluminum foil

B) Pipette (10 mL; large tip opening) and a bulb, sterilized by autoclaving at 121°C for 20 min

Preparation of mLS medium (medium no. 1)

1. Dissolve the following chemicals in approximately 800 mL of distilled water.

MS salt mix	1 bag (1 L)
Sucrose	30 g

2. Add following stock solutions, and fill up to approximately 950 mL with distilled water.

BY2_P	2.5 mL
LS_VT_modified	2.5 mL
2,4-D (0.2 mg/mL)	1 mL

3. Adjust the pH of the solution to 5.8 with KOH (1 N), and fill up to 1 L with distilled water.
4. Pour 95 mL of the medium into a 300-mL flask.
5. Autoclave the flask at 121°C for 20 min.

Methods

1. Agitate a 7-day-old culture well and transfer 3–3.2 mL of cell suspension to 95 mL of fresh mLS medium with a pipette.
2. Incubate cell cultures on a rotary shaker at 130 rpm under the dark condition at 27°C.

Notes

- For domestic customers: We send GF11 cell suspension in a 50-mL disposable conical centrifuge tube. The cells should be transferred to fresh mLS medium immediately after arrival.
- For overseas customers: We send GF11 cells placed on semi-solid mLS medium in a 250-mL disposable Erlenmeyer flask. To re-establish cell suspension cultures, refer to the culture initiation protocol.
- In order to maintain GF11 cell suspension cultures stably, it is essential to transfer an adequate amount of cells to fresh mLS medium in every subculture. The amount of cells may vary from one lab to another, because proliferation of GF11 cells is affected by culture conditions, such as a room temperature, rotation speed of a rotary shaker, and aeration condition of the culture.
- A low growth rate of the parent BY-2 cells is sometimes caused by poor aeration (Kumagai-Sano *et al.* 2007). In order to obtain good aeration of a suspension cul-

ture, a silicone sponge plug may be used instead of the aluminum foil cap (e.g., cap-type Silicosen; Shin-Etsu Polymer, Tokyo, Japan; <https://www.shinpoly.co.jp/en/product/product/medical/plugs.html>).

References

- Kumagai-Sano F, Hayashi T, Sano T, Hasezawa S (2007) Cell cycle synchronization of tobacco BY-2 cells. *Nature Protocols* 1: 2621–2627. DOI: [10.1038/nprot.2006.381](https://doi.org/10.1038/nprot.2006.381)
- Nagata T, Nemoto Y, Hasezawa S (1992) Tobacco BY-2 cell line as the “HeLa” cell in the cell biology of higher plants. *International Review of Cytology* 132: 1–30. DOI: [10.1016/S0074-7696\(08\)62452-3](https://doi.org/10.1016/S0074-7696(08)62452-3)
- Sano T, Higaki T, Oda Y, Hayashi T, Hasezawa S (2005) Appearance of actin microfilament ‘twin peaks’ in mitosis and their function in cell plate formation, as visualized in tobacco BY-2 cells expressing GFP-fimbrin. *Plant Journal* 44: 595–605. DOI: [10.1111/j.1365-313X.2005.02558.x](https://doi.org/10.1111/j.1365-313X.2005.02558.x)

Appendix A: Formulation of culture medium

Table A.1. modified Linsmaier and Skoog medium
(medium no. 1)

Chemical	Concentration (mg/L)
KNO ₃	1900
NH ₄ NO ₃	1650
CaCl ₂ ·2H ₂ O	440
MgSO ₄ ·7H ₂ O	370
KH ₂ PO ₄	370
H ₃ BO ₃	6.2
MnSO ₄ ·4H ₂ O	22.3
ZnSO ₄ ·7H ₂ O	8.6
KI	0.83
Na ₂ MoO ₄ ·2H ₂ O	0.25
CuSO ₄ ·5H ₂ O	0.025
CoCl ₂ ·6H ₂ O	0.025
FeSO ₄ ·7H ₂ O	27.8
Na ₂ -EDTA	37.3
Thiamine·HCl	1
<i>myo</i> -Inositol	100
Sucrose	30000
2,4-D sodium monohydrate	0.236