

rpc00044

Nicotiana glauca G89 callus culture

Components

- A 9-cm plastic Petri dish, containing cells placed on semi-solid medium

Notice

- Subculture the cells to fresh medium immediately after arrival [[Notes I](#)].
- Do not store the cell culture in a refrigerator and a freezer.
- Maintain aseptic conditions of the cell culture, and work in a laminar flow cabinet.

Method

- Culture medium: MS medium, 1 mg/L IAA, 0.9% (w/v) agar, pH 5.7 (medium no. 35) [[Materials III](#)]
- Culture conditions: 27°C, dark [[Methods II](#)]
- Subculture: 28-day intervals [[Methods I](#)]

Citation of cell line

When results obtained by using this cell line are published in a scientific journal, it should be cited in the following manner: “*Nicotiana glauca* G89 cell line (rpc00044) was provided by the RIKEN BRC through the National BioResource Project of the MEXT, Japan.”

Experimental Plant Division
RIKEN BioResource Research Center (BRC)
Koyadai 3-1-1, Tsukuba, Ibaraki 305-0074
Japan
FAX: +81 29 836 9053
E-mail: plant.brc@riken.jp
<http://epd.brc.riken.jp/en/>

Introduction

Tobacco G89 cell line was established from a seedling stem of *Nicotiana glauca* Graham (Syōno and Fujita 1994). The G89 cells are grown on a Murashige and Skoog (MS) medium supplemented with 1 mg/L indole-3-acetic acid (IAA), and solidified with 0.9% (w/v) agar, pH 5.7. Our G89 cell culture has been maintained in the dark at 27°C and subcultured at 28-day intervals.

Materials

Chemicals and stock solutions

(All stock solutions are stored at 4°C)

A) MS salt mix

Murashige and Skoog Plant Salt Mixture, FUJIFILM Wako Pure Chemical Corporation (#392-00591)

B) Sucrose

C) MS_VT

Nicotinic acid	0.5 mg/mL
Pyridoxine·HCl	0.5 mg/mL
Thiamine·HCl	0.1 mg/mL
Glycine	2 mg/mL

D) MS_inositol

<i>myo</i> -Inositol	40 mg/mL
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E) IAA (1 mg/mL)

IAA·K	1.217 mg/mL
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Potassium 3-indoleacetate, FUJIFILM Wako Pure Chemical Corporation (#160-07531)

F) Agar, powder

G) KOH (1 N)

Glassware and equipment

A) Erlenmeyer flask (100 mL), capped with two layers of aluminum foil

B) Forceps, sterilized before use

Preparation of MS medium (medium no. 35)

1. Dissolve the following chemicals in approximately 800 mL of distilled water.

MS salt mix	1 bag (1 L)
Sucrose	30 g

2. Add following stock solutions, and fill up to approximately 950 mL with distilled water.

MS_VT	1 mL
MS_inositol	2.5 mL
IAA (1 mg/mL)	1 mL

3. Adjust the pH of the solution to 5.7 with KOH (1 N), and fill up to 1 L with distilled water.
4. Pour 40 mL of the medium into a 100-mL flask containing 0.36 g of agar.
5. Autoclave the flask at 121°C for 20 min.

Methods

1. Pick up an appropriate amount of callus cells from a 28-day-old culture with a forceps and place the cells onto fresh MS medium.
2. Incubate cell cultures under the dark condition at 27°C.

Notes

- We send G89 cells on semi-solid MS medium in a 9-cm disposable Petri dish. The cells should be subcultured to fresh MS medium immediately after arrival.
- In order to maintain G89 callus culture stably, it is essential to observe the growth of cells carefully. Because proliferation of G89 cells is affected by culture conditions, such as a room temperature, aeration conditions of the culture and so on, an amount of cells transferred to fresh medium and the subculture intervals may vary from one lab to another. We usually inoculate three pieces of G89 callus (about 5-mm in diameter) on 40 mL of MS medium in a 100-mL flask, and culture them for 28 days.

References

- Syōno K, Fujita T (1994) Habituation as a tumorous state that is interchangeable with a normal state in plant cells. *International Review of Cytology* 152: 265–299. DOI: [10.1016/S0074-7696\(08\)62559-0](https://doi.org/10.1016/S0074-7696(08)62559-0)

Appendix A: Formulation of culture medium

Table A.1. Murashige and Skoog medium
(medium no. 35)

Chemical	Concentration (mg/L)
KNO ₃	1900
NH ₄ NO ₃	1650
CaCl ₂ ·2H ₂ O	440
MgSO ₄ ·7H ₂ O	370
KH ₂ PO ₄	170
H ₃ BO ₃	6.2
MnSO ₄ ·4H ₂ O	22.3
ZnSO ₄ ·7H ₂ O	8.6
KI	0.83
Na ₂ MoO ₄ ·2H ₂ O	0.25
CuSO ₄ ·5H ₂ O	0.025
CoCl ₂ ·6H ₂ O	0.025
FeSO ₄ ·7H ₂ O	27.8
Na ₂ -EDTA	37.3
Nicotinic acid	0.5
Pyridoxine·HCl	0.5
Thiamine·HCl	0.1
Glycine	2
<i>myo</i> -Inositol	100
Sucrose	30000
IAA·K	1.217
Agar	9000